

Glutamate (excitatory amino acid) transporters

Overview: Plasma membrane located glutamate transporters (nomenclature proposed by Amara and Arriza, 1993) are members of the solute carrier family 1 (SLC1) of sodium-dependent transporters that also includes the neutral amino acid transporters ASCT1 and ASCT2 (Palacín *et al.*, 1998; Kanai and Hediger, 2003; 2004; Beart and O'Shea, 2007). Glutamate transporters present the unusual structural motif of 8TM segments and 2 re-entrant loops (Grunwald and Kanner, 2000). The crystal structure of a glutamate transporter homologue (Glt_{ph}) from *Pyrococcus horikoshii* supports this topology and indicates that the transporter assembles as a trimer, where each monomer is a functional unit capable of substrate permeation (Yernool *et al.*, 2004; Boudker *et al.*, 2007). These structural data are in agreement with the proposed quaternary structure for EAAT2 (Gendreau *et al.*, 2004) and several functional studies that propose the monomer is the functional unit (Ryan *et al.*, 2004; Grever *et al.*, 2005; Koch *et al.*, 2007; Leary *et al.*, 2007). Splice variants of EAAT2 have recently been shown to form homomeric and heteromeric assemblies at the cell surface (Peacey *et al.*, 2009). The activity of glutamate transporters located upon both neurones (predominantly EAAT3, 4 and 5) and glia (predominantly EAAT 1 and 2) serves, dependent upon their location, to regulate excitatory neurotransmission, maintain low ambient extracellular concentrations of glutamate (protecting against excitotoxicity) and provide glutamate for metabolism including the glutamate-glutamine cycle. The Na⁺/K⁺/ATPase that drives transport has been demonstrated to co-assemble with EAAT1 and EAAT2 (Rose *et al.*, 2009). Recent evidence supports novel roles in brain for splice variants of EAAT1 and EAAT2 (Macnab and Pow, 2007; Sullivan *et al.*, 2007). Enhanced expression of EAAT2 resulting from administration of β-lactam antibiotics (e.g. ceftriaxone), or the neuroimmunophilin GPI-1046, is neuro-protective (Rothstein *et al.*, 2005; Ganel *et al.*, 2006). Enhanced expression by ceftriaxone has been proposed to occur through NF-κappaB-mediated EAAT2 promoter activation (Lee *et al.*, 2008), although the protective affects of ceftriaxone in a mouse model of multiple sclerosis are thought to be mediated by a reduction in T cell activation with no effect on EAAT2 expression, or function (Melzer *et al.*, 2008). A thermodynamically uncoupled Cl⁻ flux, activated by Na⁺ and glutamate (Kanner and Borre, 2002; Kanai and Hediger, 2003; Grever and Rauen, 2005) (or aspartate in the case of Glt_{ph}, Ryan and Mindell, 2007), is sufficiently large, in the instances of EAAT4 and EAAT5, to influence neuronal excitability (Veruki *et al.*, 2006). In the kidney, EAAT3 located in the apical membrane of proximal tubular cells is responsible for the reabsorption of glutamate (Hediger, 1999). Three structurally and functionally distinct vesicular glutamate transporters (VGLUT1, 2 and 3) of the SLC17 family are responsible for concentrating glutamate within synaptic vesicles (Reimer and Edwards, 2004).

Nomenclature	EAAT1	EAAT2	EAAT3
Other names	GLAST, SLC1A3	GLT1, SLC1A2	EAAC1, SLC1A1
Ensembl ID	ENSG000000079215	ENSG00000110436	ENSG00000106688
Endogenous substrates	L-glutamate, L-aspartate	L-glutamate, L-aspartate	L-glutamate, L-aspartate
Inhibitors (K_B or K_i)	UCPH-101 (IC_{50} = 120 nM – membrane potential assay, Jensen <i>et al.</i> (2009), DL-TBOA (9 μM)	WAY-213613 (IC_{50} = 130 nM), DL-TBOA (0.12 μM), (2S,4R)-4-methylglutamate (3.4 μM), dihydrokainate (9 μM), Threo-3-methylglutamate (18 μM)	NBI-59159 (IC_{50} = 25 nM), DL-TBOA (IC_{50} = 8 μM), L-β-BA (IC_{50} = 0.8 μM – [³ H]-D-aspartate uptake assay)
Probes	[³ H]-ETB-TBOA (K_D = 15.5 nM), [³ H]-[(2S,4R)-4-methylglutamate, [³ H]-D-aspartate, [³ H]-L-aspartate	[³ H]-ETB-TBOA (K_D = 16.2 nM), [³ H]-[(2S,4R)-4-methylglutamate, [³ H]-D-aspartate, [³ H]-L-aspartate	[³ H]-ETB-TBOA (K_D = 320 nM), [³ H]-D-aspartate, [³ H]-L-aspartate
Stoichiometry	–	3Na ⁺ : 1H ⁺ : 1glutamate (in): 1K ⁺ (out)	3Na ⁺ : 1H ⁺ : 1glutamate (in): 1K ⁺ (out)

Nomenclature	EAAT4	EAAT5
Other names	SLC1A6	SLC1A7
Ensembl ID	ENSG00000105143	ENSG00000162383
Endogenous substrates	L-glutamate, L-aspartate	L-glutamate, L-aspartate
Inhibitors (K_B or K_i)	DL-TBOA (4.4 μM), Threo-3-methylglutamate (50 μM)	DL-TBOA (3.2 μM)
Probes	[³ H]-ETB-TBOA (K_D = 24.8 nM), [³ H]-D-aspartate, [³ H]-L-aspartate	[³ H]-ETB-TBOA (K_D = 29.5 nM), [³ H]-D-aspartate, [³ H]-L-aspartate

The K_B (or K_i) values reported, unless indicated otherwise, are derived from transporter currents mediated by EAATs expressed in voltage-clamped *Xenopus laevis* oocytes (Vandenberg *et al.*, 1997; Shimamoto *et al.*, 1998; Eliasof *et al.*, 2001; Shigeri *et al.*, 2001). K_B (or K_i) values derived in uptake assays are generally higher (e.g. Shimamoto *et al.*, 1998). In addition to acting as a poorly transportable inhibitor of EAAT2, (2S,4R)-4-methylglutamate, also known as SYM2081, is a competitive substrate for EAAT1 (K_M = 54 μM; Vandenberg *et al.*, 1997; Huang *et al.*, 2009) and additionally is a potent kainate receptor agonist (Zhou *et al.*, 1997), which renders the compound unsuitable for autoradiographic localisation of EAATs (Apricò *et al.*, 2007). Similarly, at concentrations that inhibit EAAT2, dihydrokainate binds to kainate receptors (Shimamoto *et al.*, 1998). WAY-855 and WAY-213613 are both non-substrate inhibitors with a preference for EAAT2 over EAAT3 and EAAT1 (Dunlop *et al.*, 2003; Dunlop *et al.*, 2005). NBI-59159 is a non-substrate inhibitor with modest selectivity for EAAT3 over EAAT1 (>10-fold) and EAAT2 (fivefold) (Coon *et al.*, 2004; Dunlop, 2006). Analogously, L-β-threo-benzyl-aspartate (L-β-BA) is a competitive non-substrate inhibitor that preferentially blocks EAAT3 versus EAAT1, or EAAT2 (Esslinger *et al.*, 2005). [³H]-[(2S,4R)-4-methylglutamate demonstrates low affinity binding (K_D ≅ 6.0 μM) to EAAT1 and EAAT2 in rat brain homogenates (Apricò *et al.*, 2001) and EAAT1 in murine astrocyte membranes (Apricò *et al.*, 2004), whereas [³H]-ETB-TBOA binds with high affinity to all EAATs other than EAAT3 (Shimamoto *et al.*, 2007). The novel isoxazole derivative (-)-HIP-A may interact at the same site as TBOA and preferentially inhibit reverse transport of glutamate (Colleoni *et al.*, 2008). Threo-3-methylglutamate induces substrate-like currents at EAAT4, but does not elicit heteroexchange of [³H]-aspartate in synaptosome preparations, inconsistent with the behaviour of a substrate inhibitor (Eliasof *et al.*, 2001). Parawixin1, a compound isolated from the venom from the spider *Parawixia bistriata* is a selective enhancer of the glutamate uptake through EAAT2 but not through EAAT1 or EAAT3 (Fontana *et al.*, 2003; 2007). In addition to the agents listed in the table, DL-threo-β-hydroxyaspartate and L-trans-2,4-pyrrolidine dicarboxylate act as non-selective competitive substrate

inhibitors of all EAATs. Zn^{2+} and arachidonic acid are putative endogenous modulators of EAATs with actions that differ across transporter subtypes (reviewed by Vandenberg et al., 2004).

Abbreviations: DL-TBOA, DL-threo- β -benzyloxyaspartate; GPI-1046, (3-(3-pyridyl)-1-propyl (2S)-1-(3,3-dimethyl-1,2-dioxopentyl)-2-pyrrolidinecarboxylate; ETB-TBOA, (2S, 3S)-3-[3-[4-ethylbenzoylamino]benzyloxy]aspartate; (-)-HIP-A, (-)-3-hydroxy-4,5,6,6a-tetrahydro-3aH-pyrrolo[3,4-d]-isoxazole-4-carboxylic acid; NBI-59159 (also known as WAY-209429), (N-4-(9H-fluoren-2-yl)-L-asparagine; WAY-855, 3-amino-tricyclo[2.2.1.0^{2,6}]heptane-1,3-dicarboxylic acid; WAY-213613, N(4)-[4-(2-bromo-4,5-difluorophenoxy)phenyl]-L-asparagine; UCPH-101, 2-amino-4-(4-methoxyphenyl)-7-(naphthalene-1-yl)-5-oxo-5,6,7,8-tetrahydro-4H-chromene-3-carbonitrile

Further Reading

- Beart PM, O'Shea RD (2007). Transporters for L-glutamate: an update on their molecular pharmacology and pathological involvement. *Br J Pharmacol* 150: 5–17.
- Bridges RJ, Esslinger CS (2005). The excitatory amino acid transporters: pharmacological insights on substrate and inhibitor specificity of the EAAT subtypes. *Pharmacol Ther* 107: 271–285.
- Bunch L, Erichsen MN, Jensen AA (2009). Excitatory amino acid transporters as potential drug targets. *Expert Opin Ther Targets* 13: 719–731.
- Danbolt NC (2001). Glutamate uptake. *Prog Neurobiol* 65: 1–105.
- Dunlop J (2006). Glutamate based therapeutic approaches: targeting the glutamate transport system. *Curr Opin Pharmacol* 6: 103–107.
- Dunlop J, Butera JA (2006). Ligands targeting the excitatory amino acid transporters (EAATs). *Curr Top Med Chem* 6: 1897–1906.
- Grewer C, Gameiro A, Zhang Z, Tao Z, Braams S, Rauen T (2008). Glutamate forward and reverse transport: from molecular mechanism to transporter-mediated release after ischemia. *IUBMB Life* 60: 609–619.
- Grewer C, Rauen T (2005). Electrogenic glutamate transporters in the CNS: molecular mechanism, pre-steady-state kinetics, and their impact on synaptic signaling. *J Membr Biol* 203: 1–20.
- Hediger MA (1999). Glutamate transporters in kidney and brain. *Am J Physiol* 277: F487–F492.
- Hinoi E, Takarada T, Tsuchihashi Y, Yoneda Y (2005). Glutamate transporters as drug targets. *Curr Drug Targets CNS Neurol Disord* 4: 211–220.
- Huang YH, Bergles DE (2004). Glutamate transporters bring competition to the synapse. *Curr Opin Neurobiol* 14: 346–352.
- Kanai Y, Hediger MA (2003). The glutamate and neutral amino acid transporter family: physiological and pharmacological implications. *Eur J Pharmacol* 479: 237–247.
- Kanai Y, Hediger MA (2004). The glutamate/neutral amino acid transporter family SLC1: molecular, physiological and pharmacological aspects. *Pflügers Archiv* 447: 465–468.
- Kanner BI, Borre L (2002). The dual-function glutamate transporters: structure and molecular characterisation of the substrate-binding sites. *Biochim Biophys Acta* 1555: 92–95.
- Kanner BI (2006). Structure and function of sodium-coupled GABA and glutamate transporters. *J Membr Biol* 213: 89–100.
- Kanner BI, Zomot E (2008). Sodium-coupled neurotransmitter transporters. *Chem Rev* 108: 1654–1668.
- Palacín M, Estévez R, Bertran J, Zorano A (1998). Molecular biology of mammalian plasma membrane amino acid transporters. *Physiol Rev* 78: 969–1054.
- Reimer RJ, Edwards RH (2004). Organic anion transport is the primary function of the SLC17/type I phosphate transporter family. *Pflügers Archiv* 447: 629–635.
- Ryan RM, Vandenberg RJ (2005). A channel in a transporter. *Clin Exp Pharmacol Physiol* 32: 1–6.
- Sheldon AL, Robinson MB (2007). The role of glutamate transporters in neurodegenerative diseases and potential opportunities for intervention. *Neurochem Int* 51: 333–355.
- Shigeri Y, Seal RP, Shimamoto K (2004). Molecular pharmacology of glutamate transporters, EAATs and VGLUTs. *Brain Res Brain Res Rev* 45: 250–265.
- Shimamoto K (2008). Glutamate transporter blockers for elucidation of the function of excitatory neurotransmission systems. *Chem Rec* 8: 182–199.
- Sonders MS, Quick M, Javitch JA (2005). How did the neurotransmitter cross the bilayer? A closer look. *Curr Opin Neurobiol* 15: 296–304.
- Tzingouris AV, Wadiche JI (2007). Glutamate transporters: confining runaway excitation by shaping synaptic transmission. *Nat Rev Neurosci* 8: 935–947.
- Vandenberg RJ (2006). Mutational analysis of glutamate transporters. *Handb Exp Pharmacol* 175: 113–135.
- Vandenberg RJ, Huang S, Ryan RM (2008). Slips, leaks and channels in glutamate transporters. *Channels (Austin)* 2: 51–58.
- Vandenberg RJ, Ju P, Aubrey KR, Ryan RM, Mitrovic AD (2004). Allosteric modulation of neurotransmitter transporters at excitatory synapses. *Eur J Pharm Sci* 23: 1–11.
- Vandenberg RJ, Ryan RM (2005). How and why are channels in transporters? *Sci STKE* 2005(280): pe17.

References

- Amara SG, Arriza JL (1993). *Curr Opin Neurobiol* 3: 337–344.
- Apricò K et al. (2001). *J Neurochem* 77: 1218–1225.
- Apricò K et al. (2004). *J Neurosci Res* 75: 751–759.
- Apricò K et al. (2007). *Neurochem Int* 51: 507–516.
- Boudker O et al. (2007). *Nature* 445: 387–393.
- Colleoni S et al. (2008). *J Pharmacol Exp Ther* 326: 646–656.
- Coon TR et al. (2004). *Abstract Viewer/Itinerary Planner, Program No. 168.10*. Society for Neuroscience: Washington, DC.
- Dunlop J et al. (2003). *Br J Pharmacol* 140: 839–846.
- Dunlop J et al. (2005). *Mol Pharmacol* 68: 974–982.
- Eliasof S et al. (2001). *J Neurochem* 77: 550–557.
- Esslinger CS et al. (2005). *Neuropharmacology* 49: 850–861.
- Fontana AC et al. (2003). *Br J Pharmacol* 139: 1297–1309.
- Fontana AC et al. (2007). *Mol Pharmacol* 72: 1228–1237.
- Ganel R et al. (2006). *Neurobiol Dis* 21: 556–567.
- Gendreau S et al. (2004). *J Biol Chem* 279: 39505–39512.
- Grewer C et al. (2005). *Biochemistry* 44: 11913–11923.
- Grunwald M, Kanner BI (2000). *J Biol Chem* 275: 9684–9698.
- Huang S et al. (2009). *J Biol Chem* 284: 4510–4515.
- Koch HP et al. (2007). *J Neurosci* 27: 2943–2947.
- Jensen AA et al. (2009). *J Med Chem* 52: 912–915.
- Leary GP et al. (2007). *J Neurosci* 27: 2938–2942.
- Lee SG et al. (2008). *J Biol Chem* 283: 13116–13123.
- Macnab LT, Pow DV (2007). *Neuroscience* 150: 705–711.
- Melzer N et al. (2008). *PLoS ONE* 3: e3149.
- Peacey E et al. (2009). *Mol Pharmacol* 75: 1062–1073.
- Rose EM et al. (2009). *J Neurosci* 29: 8143–8155.
- Rothstein JD et al. (2005). *Nature* 433: 73–77.
- Ryan RM et al. (2004). *J Biol Chem* 279: 20742–20751.
- Ryan RM, Mindell JA (2007). *Nat Struct Mol Biol* 14: 365–371.
- Shigeri Y et al. (2001). *J Neurochem* 79: 279–302.
- Shimamoto K et al. (1998). *Mol Pharmacol* 53: 195–201.

Shimamoto K *et al.* (2007). *Mol Pharmacol* **71**: 294–302.
Sullivan SM *et al.* (2007). *Neuroscience* **149**: 434–445.
Veruki ML *et al.* (2006). *Nat Neurosci* **11**: 1388–1396.

Vandenberg RJ *et al.* (1997). *Mol Pharmacol* **51**: 809–815.
Yernool D *et al.* (2004). *Nature* **431**: 811–818.
Zhou LM *et al.* (1997). *J Pharmacol Exp Ther* **280**: 422–427.